



## A medical bioscience study of serum levels of paraoxonase 1 activity and total antioxidant capacity relationship with anti-mullerian hormone in polycystic ovary syndrome

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### Abstract

The medical bioscience aims at training and developing scientists who will conduct research with a view to reduce the many lifestyle associated sicknesses. Polycystic ovary disease (PCOS) is a multifactorial disease and its prevalence rate increased significantly in the last few years. It is considered a cause of infertility and delayed reproduction, in addition to the many complications associated with the disease, which affect the lives of the patients in many aspects. During the study of the most important factors related to disease it is clear that the rise in AMH is indicative of degree of severity of disease, where it reflects the number of small follicles in the ovary. Hyperinsulinemia and insulin resistance with hyper- androgenemia conditions associated with PCOS enters the patients in vicious circle of factors where there is an increase in oxidative stress and decrease in antioxidants, including PON-1 and TAOC as the results showed, this in turn more increases the resistance of insulin and high level of testosterone followed by various complications on the heart and fertility and obesity and others. Serum AMH and PON-1 was measured using enzyme linked immunosorbent kit [Elisa], while FSH, LH, testosterone, prolactin and estradiol were determined by VIDAS kit method (enzyme linked fluorescent assay) and TAOC measured by colorimetric method.

**Keywords:** medical bioscience, anti-mullerian hormone (AMH), PON-1, TAOC, PCOS

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### INTRODUCTION

From the perspective of medical bioscience, polycystic ovarian syndrome is a multifactorial disorder affecting women in reproductive age, and is considered as systemic disease not only local disease exclusive in the ovary, this appellation come from its dire consequences on many systems of the females' body, since this syndrome associated with hyperinsulinemia and insulin resistance this lead to type 2 diabetes and central and/or general obesity and besides dyslipidemia in addition to fore mentioned factors all cause cardiovascular disease. Its frequently accompanies with oxidative stress, hyperandrogenemia and chronic inflammation (De Leo et al. 2016, Lim et al. 2012).

The mechanism hidden behind increased oxidative stress in PCOS not fully clear until now, there are many studied linked between increased oxidative stress in PCOS and insulin resistance that the most important feature in this disease. Oxidative stress is the condition of imbalance between oxidant and antioxidants. Antioxidants are chemical compounds that throw away, scavenge, and stifle the formation of free radicals, or

suppress their actions (Elmasry et al. 2015, Zuo et al. 2016).

The HDL-associated paraoxonase 1 (PON1) enzyme is a glycoprotein composed of 354 a.a (Komoda 2013). Paraoxonase 1 in the serum secreted by the liver consider as antioxidant enzyme and is responsible for hydrolases organophosphate pesticides and neurotoxic compound (Fuhrman 2012, Mehdi and Rizvi 2012). It's also involve in increase macrophage associated-cholesterol efflux and degrades homocysteine thiolactone thereby preventing protein modification, and stabilizes free radicals, thus maintaining membrane integrity (Chistiakov et al. 2017). Furthermore, PON1 has been reported to stimulate insulin secretion and biosynthesis and boosted uptake of glucose by increased Glucose transporter type 4 (GLUT-4) expression, decrease in pon-1 activity may associated with increased insulin resistance as it happens in

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diabetes mellitus and AHD (Koren-Gluzer et al. 2011, 2013, Mehdi and Rizvi 2012). Studies in genetic aspect indicate that some genetic variation in pon-1 gene contribute to reduced expression of it (Dadachanji et al. 2018).

Total antioxidant capacity (TAOC) is defined as the ability of serum to quench free radical production and thus protecting the cell structure from molecular damage. Studies on this marker and its relation with PCOS is conflicting, can be measured its value in various method but the most common one its spectrophotometric assay (Hyderali and Mala 2015, Jeelani et al. 2017). Anti-Mullerian hormone (AMH), a dimeric glycoprotein and member of the transforming growth factor- $\beta$  family, is produced by granulosa cells in preantral and early antral follicles within the ovaries and is correlated with ovarian reserve. In PCOS, AMH reduce sensitivity of small follicles to FSH and that lead to anovulation (Piltonen et al. 2005, Rzeszowska et al. 2016). This study was designed to evaluate the serum paraoxonase (PON1) activity and total antioxidant capacity in Iraqi women with PCOS and its correlation with Anti-mullerian hormone level and other hormone and PCOS pathogenesis.

## MATERIALS AND METHODS

This study comprised 80 woman diagnosed with PCOS according to Rotterdam criteria by specialist gynecologists, and 60 healthy woman have normal regulatory menstrual cycle as control, where women who were diagnosed with PCOS had at least two of these criteria: 1- oligo ovulation and /or anovulation 2- hyperandrogenemia clinically or biochemically, and 3- polycystic ovaries on ultrasound, their age ranged from 16 to 40 year. The blood samples were collected from Al-Shatrah hospital and Alhussien teaching hospital in Thi-Qar governorate/Iraq during the period from April-September/2018. We excluded the patients with endocrine disease and those with hormonal therapy that effect on the hormones level and also excluded pregnant and smoking woman.

Blood samples (5 ml) was collected during the follicular phase (2-3) day of menstrual cycle) for both groups.

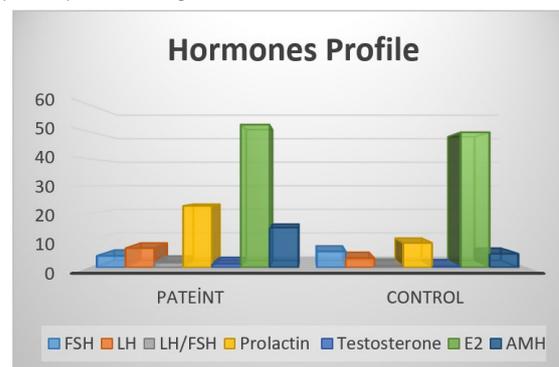
Body Mass Index (BMI) was determined by the following equation;  $BMI = \text{weight kg} / \text{height m}^2$  for both patient and control. Measure waist circumference in centimeters, measured at the smallest circumference of the waist, above the belly button and the hip circumference measured at the widest part of the buttocks, then calculated the WHR by dividing waist/hip.

PON-1 enzyme level and AMH were measured using Enzyme linked immunosorbent assay (ELISA) method, [Elabscience Biotechnology Inc and Anshlab, (both of USA origin) respectively]. While serum total antioxidant capacity was estimated by Colorimetric method

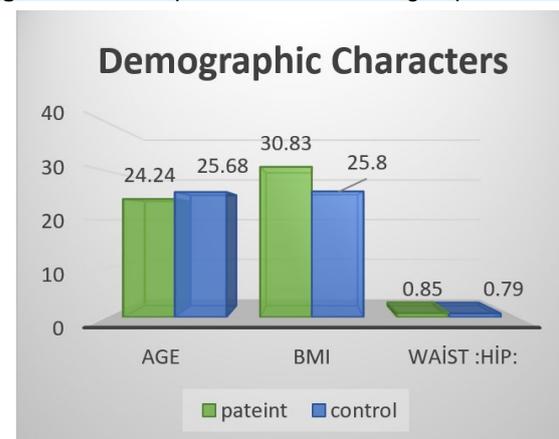
**Table 1.** Demographic features with hormonal levels in patients and control

Characteristics	Samples		P value
	Patient (80) Mean $\pm$ SD	Control (60) Mean $\pm$ SD	
Age	24.24 $\pm$ 5.78	25.68 $\pm$ 6.30	0.161 NS
BMI(k/m <sup>2</sup> )	30.83 $\pm$ 5.50	25.80 $\pm$ 4.22	0.0001**
Waist :hip:	0.85 $\pm$ 0.06	0.79 $\pm$ 0.040	0.0001**
FSH (mIU/ ml)	4.25 $\pm$ 1.58	5.90 $\pm$ 1.50	0.0001**
LH (mIU/ ml)	7.09 $\pm$ 4.37	3.37 $\pm$ 1.00	0.0001**
LH/FSH	1.84 $\pm$ 1.56	0.58 $\pm$ 0.165	0.0001**
Prolactin (ng/ml)	22.63 $\pm$ 11.80	9.02 $\pm$ 4.10	0.0001**
Testosterone(ng/ml)	1.30 $\pm$ 0.65	0.42 $\pm$ 0.12	0.0001**
E2 (Pg/ml)	52.49 $\pm$ 23.33	49.65 $\pm$ 24.80	0.489 NS
AMH(ng/ml)	14.65 $\pm$ 4.14	5.02 $\pm$ 2.02	0.0001**

\*\* (P<0.01), NS: Non-Significant.



**Fig. 1.** Hormones profile for the studied groups



**Fig. 2.** Demographical Properties for the studied groups

(Elabscience Co.) and serum level of FSH, LH, Testosterone, prolactin and E2 were determined by VIDAS which is full automated technique using principle of enzyme linked Fluorescent Assay.

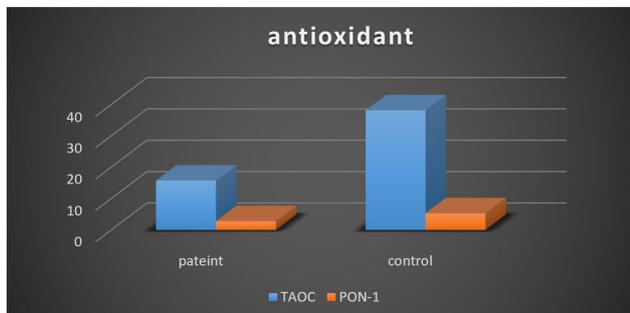
## RESULT

The data in **Table 1** showed that there was no significant difference in the mean of age in women with PCOS and control, while there was a significant difference in the BMI between those groups. Additionally, table revealed that there was a significant difference in the mean of hormones (FSH, LH, FSH/LH, PRL, testosterone, AMH) between patients and control, while the value of E2 did not reach to the statistically significant level (**Figs. 1 & 2**).

**Table 2.** Serum antioxidants level in patient and control

	Samples		P value
	Patient (80) Mean ± SD	Control (60) Mean ± SD	
TAOC(U/mL)	15.77 ± 8.34	38.16 ± 11.96	0.0001**
PON-1( ng/mL)	2.79 ± 0.82	5.33 ± 1.58	0.0001**

\*\* (P<0.01)



**Fig. 3.** Concentration of Antioxidants in patients and controls

**Table 4.** Correlation coefficient between PON-1 and other parameters

Parameters	Correlation coefficient-r and PON-1	P value
FSH	-0.030	NS
LH	-0.202	NS
LH/FSH	-0.083	NS
Prolactin	-0.102	NS
Testosterone	-0.537	**
AMH	-0.421	**
E2	-0.079	NS
TAOC	0.564	**
BMI	-0.425	**

\*\* (P<0.01), NS: Non-Significant.

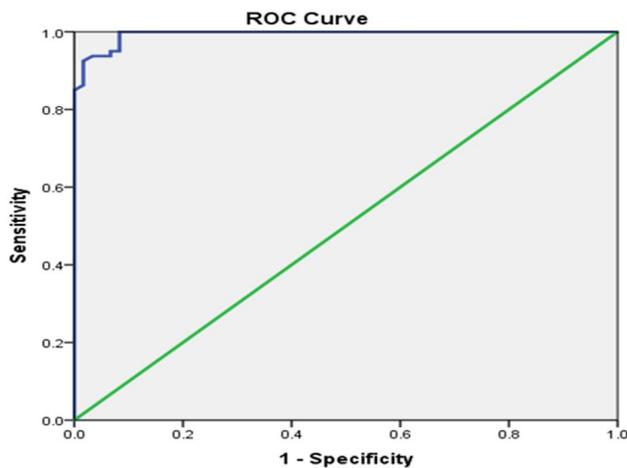
**Table 2** and **Fig. 3** illustrated that the serum level of total antioxidant capacity and paroxonase -1 were significantly decreased in the PCOS patients in comparison with control.

**Table 3** and **Fig. 4** showed the Receiver Operator Curve (ROC) Analysis, to distinguish between patient and control by using the investigated testes, the ROC analysis was utilized and help to visually understand the impact of each parameters and quantify how well each test performs by finding the area under the ROC curve. The AMH ROC curve lies to the top left corner, the greater the area underneath it. Thus, a larger area under the ROC curve implies AMH is better test for PCOS with high sensitivity and specificity (100% and 92%) at the (7.48) optimal cutoff point which give 'highest sensitivity while maintaining high specificity'. The other test also shows large area under the curve (PON-1 (AUC 0.937) and TAOC (AUC 0.945) that indicated these test have importance in comparison between PCOS woman and normal women. The sensitivity and specificity (91% and 82%) for PON-1 and (85 % and 82%) for TAOC at the optimal cutoff point.

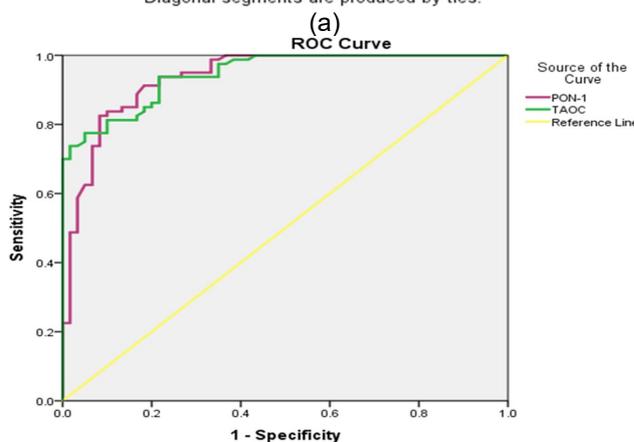
**Table 3.** (ROC) curve analysis for the AMH, PON-1 and TAOC with sensitivity and specificity

Parameters	AUC	Optimal cut off value	Sensitivity	specificity	p- value	95% Confidence Interval	
						Lower Bound	Upper Bound
AMH	0.994	7.48	100%	92%	**0.0001	0.99	1.000
PON-1	0.937	3.90	91%	82%	**0.0001	0.896	0.977
TAOC	0.945	25.34	85%	82%	**0.0001	0.913	0.978

\*\* (P<0.01)



Diagonal segments are produced by ties.



Diagonal segments are produced by ties.

(b)

**Fig. 4.** ROC curve of AMH, PON-1 and TAOC (a) ROC curve of AMH (AUC 0.994, CI 0.99-1.000) larger test result indicates more positive result. (b) ROC curve of PON-1 (AUC 0.937, CI 0.89-0.97) and TAOC (AUC 0.945, CI 0.91-0.97) smaller test result indicates more positive result all parameters have significant >0.0001

**Table 4** manifested a significant positive correlation between PON-1 (pg/ml) with TAOC (r=0.564, p<0.0001), and also noted there is significant negative correlation between PON-1 with both AMH (r=-0.421, p<0.0001) and testosterone (r=-0.537, p <0.0001).

## DISCUSSION

In polycystic ovaries, there is an increment in the number of small follicles and this enhanced the elevation in the concentration of the hormone AMH, as this hormone is excreted from the granulosa cell in these ova and this hormone has an inhibitory effect on the hormone FSH and thus prevents natural ovulation in

these patients which explained the current results (Stracquadio et al. 2018). The AUC of the serum AMH assay reached a value of 0.994 (95 % CI 0.98–1.000) with high sensitivity and specificity. This demonstrates the high diagnostic significance of AMH.

Many theories tried to explain the cause of elevation of LH in PCOS, as primary arising in LH pulse frequency (Hayes et al. 1998). But some suggested that it is due to abnormal feedback at hypothalamic level or sensitization of pituitary gland to GnRH by abnormal steroid feedback (Ehrmann 2005). Both increment in insulin and in LH, accompanied by insulin resistance, mediated high level of testosterone (Suresh and Vijayakumar 2015) Presence of hyperprolactinemia in PCOS could be clarified by many theories, one of them referred to the probable role of estrogen mainly estrone E<sub>1</sub> 'from androgenic precursors' which prompts hyperprolactinemia or increased feedback of steroid on lactotrope (Mah and Webster 2002).

Elevation in lipid profile in PCOS, an increased LDL, with a decreased HDL (Bilman and Yetik 2017, Binazir et al. 2016, Kim and Choi 2013, Kiranmayee et al. 2017),

and highness in inflammatory marker and other factors; all are reasons to oxidative stress in these patients (Alanbay et al. 2012, Blair et al. 2013). The oxidative stress status reflected by decrease in activity of PON-1 in serum of woman complaining from PCOS in addition to decrease in TAOC, the importance of these tests in explain the level of oxidative stress was further clarified by the ROC curve. The high level of oxidative stress rises the severity of PCOS disease which expressed in the high proportion of AMH, Pearson correlation demonstrate the strong correlation between AMH and pon-1 and TAOC. Moreover, increased in weight gaining or obesity of patient in turn results in increased in oxidative stress action (Azziz et al. 2016, Kheiry et al. 2013, Suresh and Vijayakumar 2015, Zuo et al. 2016).

Finally, it could be concluded that in PCOS the reduction of PON-1 and TAOC serum level is associated with the increase in the AMH hormone, which reflects the severity of the disease and also with the increase in weight, the decrease in the rate of antioxidant factors and increase the oxidative stress, as weight plays an important role in all aspect of disease.

## REFERENCES

- Alanbay I, Ercan CM, Sakinci M, Coksuer H, Ozturk M, Tapan S (2012) A macrophage activation marker chitotriosidase in women with PCOS: does low-grade chronic inflammation in PCOS relate to PCOS itself or obesity? *Archives of gynecology and obstetrics*, 286(4): 1065-71. <https://doi.org/10.1007/s00404-012-2425-0>
- Azziz R, Carmina E, Chen Z, Dunaif A, Laven JS, Legro RS, Lizneva D, Natterson-Horowitz B, Teede HJ, Yildiz BO (2016) Polycystic ovary syndrome. *Nature Reviews Disease Primers*, 2: 16057. <https://doi.org/10.1038/nrdp.2016.57>
- Bilman FB, Yetik M (2017) *Geotrichum candidum*: A rare infection agent at urinary system and review of the literature. *J Clin Exp Invest.*, 8(4): 127-9. <https://doi.org/10.5799/jcei.382434>
- Binazir MB, Alizadeh M, Nikasa P, Azhough R, Movassaghi R (2016) The Effect of a Modified World Health Organization Surgical Safety Checklist on Postoperative Complications in a Tertiary Hospital in Iran. *European Journal of General Medicine*, 13(1): 21-7. <https://doi.org/10.15197/ejgm.01442>
- Blair SA, Kyaw-Tun T, Young IS, Phelan NA, Gibney J, McEneny J (2013) Oxidative stress and inflammation in lean and obese subjects with polycystic ovary syndrome. *The Journal of reproductive medicine*, 58(3-4): 107-14.
- Chistiakov DA, Melnichenko AA, Orekhov AN, Bobryshev YV (2017) Paraoxonase and atherosclerosis-related cardiovascular diseases. *Biochimie*, 132: 19-27. <https://doi.org/10.1016/j.biochi.2016.10.010>
- Dadachanji R, Shaikh N, Patil A, Shah N, Mukherjee S (2018) PON1 promoter polymorphisms contribute to PCOS susceptibility and phenotypic outcomes in Indian women. *Gene*, 661: 34-44. <https://doi.org/10.1016/j.gene.2018.03.083>
- De Leo V, Musacchio MC, Cappelli V, Massaro MG, Morgante G, Petraglia F (2016) Genetic, hormonal and metabolic aspects of PCOS: an update. *Reproductive Biology and Endocrinology*, 14(1): 38. <https://doi.org/10.1186/s12958-016-0173-x>
- Ehrmann DA (2005) Polycystic ovary syndrome. *New England Journal of Medicine*, 352(12): 1223-36. <https://doi.org/10.1056/NEJMr041536>
- Elmasry SA, Al-Azzawi MA, Ghoneim AH, Nasr MY, AboZaid MM (2015) Role of oxidant–antioxidant imbalance in the pathogenesis of chronic obstructive pulmonary disease. *Egyptian journal of chest diseases and tuberculosis*, 64(4): 813-20. <https://doi.org/10.1016/j.ejcdt.2015.06.001>
- Fuhrman B (2012) Regulation of hepatic paraoxonase-1 expression. *Journal of lipids*. <https://doi.org/10.1155/2012/684010>

- Hayes FJ, Taylor AE, Martin KA, Hall JE (1998) Use of a gonadotropin-releasing hormone antagonist as a physiologic probe in polycystic ovary syndrome: assessment of neuroendocrine and androgen dynamics. *The Journal of Clinical Endocrinology & Metabolism*, 83(7): 2343-9. <https://doi.org/10.1210/jcem.83.7.4925>
- Hyderali BN, Mala K (2015) Oxidative stress and cardiovascular complications in polycystic ovarian syndrome. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 191:15-22. <https://doi.org/10.1016/j.ejogrb.2015.05.005>
- Jeelani H, Ganie MA, Parvez T, Fatima Q, Kawa IA, Rashid F (2017) Oxidative stress biomarkers in polycystic ovary syndrome (PCOS). *Precision medicine*, 2(1): 30-8.
- Kheiry MV, Hafezi AM, Hesarak S (2013) Bone Regeneration Using Nanotechnology–Calcium Silicate Nano-Composites. *UCT Journal of Research in Science, Engineering and Technology*, 1(1): 1-3.
- Kim JJ, Choi YM (2013) Dyslipidemia in women with polycystic ovary syndrome. *Obstetrics & gynecology science*, 56(3): 137. <https://doi.org/10.5468/ogs.2013.56.3.137>
- Kiranmayee D, Kavya K, Himabindu Y, Sriharibabu M, Madhuri GL, Venu S (2017) Correlations between anthropometry and lipid profile in women with PCOS. *Journal of human reproductive sciences*, 10(3): 167. [https://doi.org/10.4103/jhrs.JHRS\\_108\\_16](https://doi.org/10.4103/jhrs.JHRS_108_16)
- Komoda T (Editor) (2013) *The HDL handbook: biological functions and clinical implications*. Academic Press.
- Koren-Gluzer M, Aviram M, Hayek T (2013) Paraoxonase1 (PON1) reduces insulin resistance in mice fed a high-fat diet, and promotes GLUT4 overexpression in myocytes, via the IRS-1/Akt pathway. *Atherosclerosis*, 229(1): 71-8. <https://doi.org/10.1016/j.atherosclerosis.2013.03.028>
- Koren-Gluzer M, Aviram M, Meilin E, Hayek T (2011) The antioxidant HDL-associated paraoxonase-1 (PON1) attenuates diabetes development and stimulates  $\beta$ -cell insulin release. *Atherosclerosis*, 219(2): 510-8. <https://doi.org/10.1016/j.atherosclerosis.2011.07.119>
- Lim SS, Davies MJ, Norman RJ, Moran LJ (2012) Overweight, obesity and central obesity in women with polycystic ovary syndrome: a systematic review and meta-analysis. *Human reproduction update*, 18(6): 618-37. <https://doi.org/10.1093/humupd/dms030>
- Mah P, Webster J (2002) Hyperprolactinemia: etiology, diagnosis and management. *Semin Reprod Med.*: 365-74. <https://doi.org/10.1055/s-2002-36709>
- Mehdi MM, Rizvi SI (2012) Human plasma paraoxonase 1 (PON1) arylesterase activity during aging: correlation with susceptibility of LDL oxidation. *Archives of medical research*, 43(6): 438-43. <https://doi.org/10.1016/j.arcmed.2012.08.012>
- Piltonen T, Morin-Papunen L, Koivunen R, Perheentupa A, Ruokonen A, Tapanainen JS (2005) Serum anti-Müllerian hormone levels remain high until late reproductive age and decrease during metformin therapy in women with polycystic ovary syndrome. *Human Reproduction*, 20(7): 1820-6. <https://doi.org/10.1093/humrep/deh850>
- Rzeszowska M, Leszcz A, Putowski L, Hałabiś M, Tkaczuk-Włach J, Kotarski J, Polak G (2016) Anti-Müllerian hormone: structure, properties and appliance. *Ginekologia polska*, 87(9): 669-74. <https://doi.org/10.5603/GP.2016.0064>
- Stracquadanio M, Ciotta L, Palumbo MA (2018) Relationship between serum anti-Müllerian hormone and intrafollicular AMH levels in PCOS women. *Gynecological Endocrinology*, 34(3): 223-8. <https://doi.org/10.1080/09513590.2017.1381838>
- Suresh S, Vijayakumar T (2015) Correlations of insulin resistance and serum testosterone levels with LH: FSH ratio and oxidative stress in women with functional ovarian hyperandrogenism. *Indian Journal of Clinical Biochemistry*, 30(3): 345-50. <https://doi.org/10.1007/s12291-014-0447-z>
- Zuo T, Zhu M, Xu W (2016) Roles of oxidative stress in polycystic ovary syndrome and cancers. *Oxidative medicine and cellular longevity*. <https://doi.org/10.1155/2016/8589318>